

The scRNA-seq expression profiling of the receptor ACE2 and the cellular protease TMPRSS2 reveals human organs susceptible to COVID-19 infection

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Summary

Background

COVID-19 caused by SARA-CoV-2 is a disaster sweeping over 200 countries, and more than 2,150,000 people are suffering from the disease and 140,000 people died. ACE2 is a receptor protein of SARS-CoV-2, and TMPRSS2 promotes virus proliferation and transmission. Some patients developed multiple organ dysfunction syndromes other than lungs. Therefore, studying the viral susceptibility of other organs is important for a deeper understanding of viral pathogenesis.

Methods

The advantage of scRNA-seq data is the identification of cell types by clustering the gene expression of cells. ACE2 and TMPRSS2 are highly expressed in AT2 of lungs, we compared the ACE2 and TMPRSS2 expression levels of cell types from 31 organs, with AT2 of lungs to evaluate the risk of the viral infection using scRNA-seq data.

Findings

For the first time, we found the brain, gall bladder, and fallopian tube are vulnerable to COVID-19 infection. Besides, the nose, heart, small intestine, large intestine, esophagus, testis and kidney are also identified to be high-risk organs with high expression levels of ACE2 and TMPRSS2. Moreover, the susceptible organs are grouped into three risk levels based on the TMPRSS2 expression. As a result, the respiratory system, digestive system and reproductive system are at the top-risk level to COVID-19 infection.

Interpretation

This study provides evidence for COVID-19 infection in the human nervous system, digestive system, reproductive system, respiratory system, circulatory system and urinary system using scRNA-seq data, which helps for the clinical diagnosis and treatment of patients.

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Introduction

In December 2019, a novel pneumonia coronavirus, recently named Coronavirus Disease 2019 (COVID-19) by World Health Organization (WHO), was first detected in several patients in Wuhan, China. Pneumonia spread widely in more than 200 countries, and more than 2,150,000 people were suffering from the disease and over 140,000 people died, posed a major threat to the global public health. COVID-19 is caused by the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2),¹ which seriously damaged the respiratory system. Some patients developed acute respiratory infection symptoms, and even acute respiratory distress syndrome (ARDS), acute respiratory failure and other complications.^{2,3} On the other side, multiple organ dysfunction syndromes occurred in some patients and even led to death, suggesting that the virus invades other organs at the same time.²² SARS-CoV-2 enters the cell via angiotensin-converting enzyme II (ACE2), the receptor protein of SARS-CoV and NL63,⁴⁻⁹ while the cellular protease TMPRSS2 promotes the transmission during the viral infection.⁸⁻¹² It is reasonable to predict the risk of organs vulnerable to COVID-19 infection using the expression level of ACE2 and TMPRSS2.

The advantage of scRNA-seq over bulk RNA sequence is the identification of cell types by clustering the gene expression of cells. To explore the potentially susceptible organs, previous studies analyzed the ACE2 expression of some human organs using scRNA-seq data. The gross anatomical observation indicates that the lesions caused by the novel coronavirus are mainly in the lung.¹³ ACE2 is mainly expressed in type II alveolar cells (AT2) in the lung,¹⁴ which implies AT2 cell is the cell type vulnerable to COVID-19 infection. Many researchers have obtained the susceptibility of other organs from different system using ACE2 expression in AT2 cells as a reference. Following this way, many respiratory organs were considered, and ACE2 was reported to be highly expressed in the nasal tissue, mouth, airway, and lung.¹⁵⁻¹⁷ The esophagus, large intestine (ileum and colon), and pancreas were identified as high-risk organs in the digestive system by the following papers.¹⁶⁻¹⁹ The kidney and bladder as major organs of urinary system were also indicated to be high ACE2-expressed.^{17,20,21} Besides, the testes and uterus were manifested to be susceptible organs, implying that the reproductive system was a potential route of viral infection.^{22,23}

In this paper, the expression level of ACE2 and TMPRSS2 in different cell clusters of organs from nine major systems (including the respiratory system, digestive system, nervous system, endocrine system, reproductive system, circulatory system, urinary system, and motor systems) were obtained using the scRNA-seq data. For the first time, we found the brain (substantia nigra and cortex), fallopian tube, and gall bladder are vulnerable to COVID-19 infection; Besides, the lung, nose (nasal brushing epithelial cells, nasal turbinate epithelial cells, and nasal airway epithelial cells), heart, small intestine (jejunum, ileum, and duodenum), large intestine (rectum and colon), esophagus, testes, and kidney are predicted as high-risk organs under a more rigorous standard; Moreover, as the spike (S) protein initiated by TMPRSS2 is essential for the entry of the virus into the target cells and the transmission of the virus in the infected host, we used the expression level of TMPRSS2 to predict the risk level of each susceptible organ, and found that the respiratory system, digestive system, and reproductive system are the top level vulnerable to COVID-19 infection.

Methods

The scRNA-seq data of healthy human available are collected for the analysis, including 31 organs from nine major human systems (Table 1), and the details of the data resources are in Supplementary File Table S1.

Table 1: Human organs available for scRNA-seq data analysis

Human systems	Human organs	Human systems	Human organs
Digestive system	Esophagus ²⁴	Respiratory system	Nose (Nasal brushing epithelial cells, Nasal turbinate epithelial cells, and Nasal airway epithelium) ³²
	Small intestine (Jejunum, Ileum, and Duodenum) ²⁵		Bronchus ³²
	Large intestine (Rectum, Colon)		Lung ³³
	Stomach ²⁵		Trachea ²⁵
	Liver ²⁵	Circulatory system	Peripheral blood ²⁵
	Gall bladder ²⁵		Heart ³⁴
	Pancreatic islets ²⁶		Artery ²⁵
			Spleen ²⁴
Nervous system	Brain (Substantia nigra and cortex, Neuronal Epithelium, ²⁷ and Hippocampus ²⁸)	Urinary system	Kidney ²⁵
	Cerebellum ²⁵		Ureter ²⁵
	Spinal cord ²⁵		Prostate ²⁵
		Immune system	Tonsil (Tonsil dendritic cells) ³⁵
Reproductive system	Ovary ²⁹		Bone marrow ²⁵
	Fallopian tube ³⁰		Lymph nodes ³⁶
	Uterus ²⁵		Thyroid ²⁵
	Testis ³¹		Thymus ²⁵
Motor system	Muscle ²⁵	Endocrine system	

R package, Seurat, was applied to analyze the scRNA-seq data (details see Supplementary File Text S1). The expression level of ACE2 in AT2 cells is the primary reference to infer the susceptibility of other organs and tissues. The ratio of ACE2-expressed cells over total AT2 cells is 0.79% which is the mean of 8 samples. To determine the susceptibility of the organs, the expression level of TMPRSS2 is another reference. The cell cluster, in which TMPRSS2 is expressed and ACE2-expressed cell ratio is equal to or greater than AT2 cells at the same time, is identified as highly susceptible cells, and the corresponding organs are vulnerable to COVID-19.

As the TMPRSS2 is a key protease to help the SARS-CoV-2 enter the target cells, and the expression level of TMPRSS2 implies the sensitivity of the cell to COVID-19 infection. Therefore, the susceptible organs were sorted into three groups by the ratios of TMPRSS2-expressed cells. In details, the cells with TMPRSS2-expressed ratio greater 20% are defined as level 1, which is at the highest risk; the cells with ratio more than 5% and less than 20% are level 2, which means a higher risk; the rest are level 3, an existing risk of infection.

Role of the funding source

The funder of the study had no role in study design, data collection, data analysis, data interpretation, or writing of the report. The corresponding author had full access to all the data in the study and had final responsibility for the decision to submit for publication.

Results

Respiratory System

The scRNA-seq data of the lung, nose, trachea, and bronchus in the respiratory system were collected for the analysis. In the lung, AT2 cells contain an average of 0.79% ACE2-expressed cells across 8 samples (Fig. 1a, Fig. 1b, Supplementary File: Figure S1-S8, Table 2), and the expression levels of ACE2 and TMPRSS2 are high in AT2 cells (Fig. 1c, Fig. 1d). The data of nose (nasal brushing epithelial cells, nasal turbinate epithelial cells, and nasal airway epithelial cells) contains ACE2-expressed and TMPRSS2-expressed cell clusters (Supplementary File: Figure S9-S11), and the ratios of ACE2-expressed cells of these cell clusters are all over 0.79% (Table 2), thus the nose is identified as the high-risk organ. Low ratio of ACE2-expressed cells in the bronchus and trachea means they are low-risk infection organs (Supplementary File: Figures S12-S13).

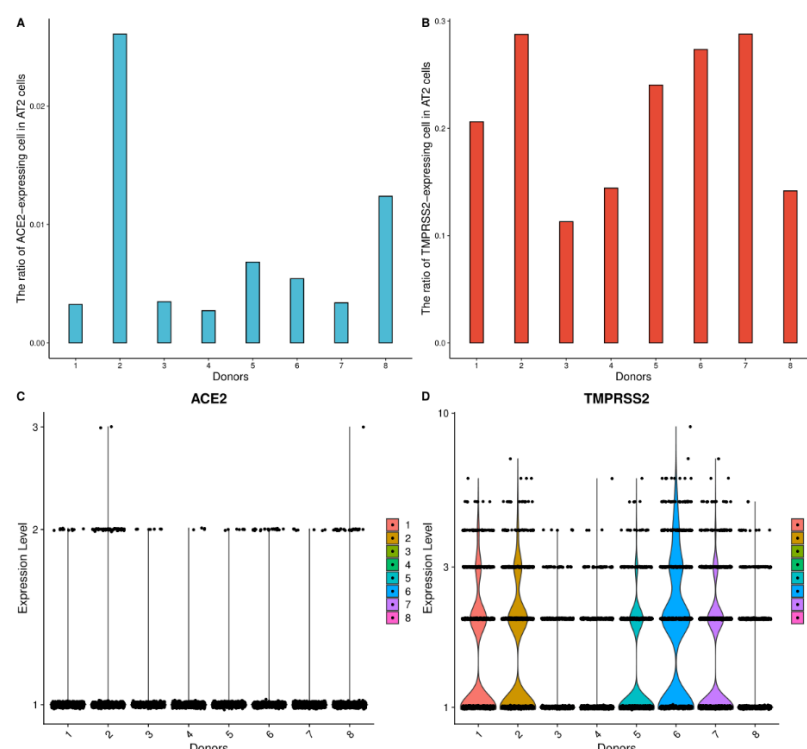


Fig. 1 High ACE2 and TMPRSS2 expression level of AT2 cells in the lung. a) The ratio of ACE2-expressed cells in AT2 cells from 8 samples. b) The ratio of TMPRSS2-expressed cells in AT2 cells from 8 samples. c) The expression distribution of ACE2 in AT2 cells across 8 samples. d) The expression distribution of TMPRSS2 in AT2 cells across 8 samples.

Table 2: Cell types with high expression levels of ACE2 and TMPRSS2 in respiratory system

Organs	Cell type	Ratio of ACE2-expressed cells	Ratio of TMPRSS2-expressed cells
Lung	AT2 cells	0.79%	21.2%
Nose (Nasal turbinate epithelial cells)	Mesenchymal stromal cells	1.7%	8.5%
	Plasma cells	2.0%	8.8%
Nose (Nasal brushing epithelial cells)	Plasma cells	4%	13.8%
Nose (Nasal airway epithelial cells)	Nasal airway epithelial cells	8.4%	23.1%

Digestive system

The scRNA-seq data of jejunum, ileum, duodenum, rectum, colon, esophagus, gall bladder, pancreatic islets, liver, and stomach from the digestive system were collected for the analysis. The primordium cells from the gall bladder contain 2 6% TMPRSS2-expressed cells and 2 2% ACE2-expressed cells (Fig. 2, Table 3), which means the gall bladder is vulnerable to the COVID-19 infection. Moreover, small intestine (jejunum, ileum, and duodenum), large intestine (rectum and colon), and esophagus are identified as high-risk organs (Supplementary File: Figures S14–S19, Table 3). However, no cell clusters from the liver, stomach, and pancreatic islets data show high ACE2 and TMPRSS2 expression level (Supplementary File: Figures S20–S22), which demonstrates a low infection risk.

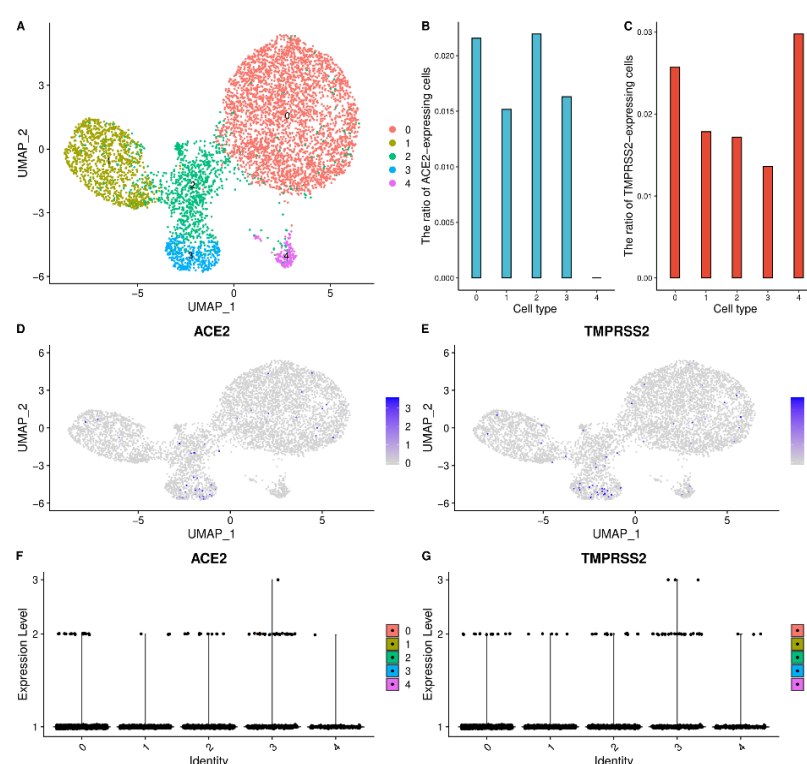


Fig. 2 High ACE2 and TMPRSS2 expression level of the primordium cells in the gall bladder. a) UMAP visualization of clustering results for gall bladder cells. b) The ratio of ACE2-expressed cells in each cell cluster. c) The ratio of TMPRSS2-expressed cells in each cell cluster. d) ACE2 expression level in each cell cluster on the UMAP plot. e) TMPRSS2 expression level in each cell cluster on the UMAP plot. f) The expression distribution of ACE2 across each cell cluster. g) The expression distribution of TMPRSS2 across each cell cluster.

Table 3: Cell types with high expression levels of ACE2 and TMPRSS2 in digestive system

Organs	Cell type	Ratio of ACE2-expressed cells	Ratio of TMPRSS2-expressed cells
Gall bladder	Primordium cell	2.2%	2.6%
Small intestine (Jejunum)	Enterocyte progenitor cell	14.0%	6.6%
	Goblet cell	9.1%	2.6%
Small intestine (Ileum)	Intestinal epithelial stem cell	1.5%	4.2%
	Enterocyte progenitor cell	2.4%	3.6%
Small intestine (Duodenum)	LGR5+ stem cell	5.2%	4.5%
	Intestinal epithelial stem cell	3.9%	5.8%
	Enterocyte progenitor cell	4.4%	6.8%
	Tuft progenitor cell	7.4%	6.6%
	Enteroendocrine cell	9.4%	3.8%
Large intestine (Rectum)	Goblet progenitor cell	2.8%	62.5%
	MKI67+ progenitor cell	10.3%	75.4%
	Enterocyte	13.2%	77.5%
	Goblet cell	18.2%	87.3%
Large intestine (Colon)	Enterocyte	5.7%	52.1%
	Goblet cell	6.7%	52.4%
Esophagus	Secretory progenitor cell	1.5%	5.0%

Nervous System

The scRNA-seq of substantia nigra and cortex, hippocampus, cerebellum, spinal cord, and neuronal epithelium from the nervous system were collected to infer the susceptibility of the organs. The analysis results show that ACE2 is expressed in the oligodendrocyte precursor cells and astrocytes of the substantia nigra and cortex with a high level, and TMPRSS2 is expressed as well. In details, astrocytes contain 1.9% ACE2-expressed cells and oligodendrocyte precursor cells contain 1.6% ACE2-expressed cells (Fig. 3, Table 4). Therefore, the substantia nigra and cortex are predicted as high-risk tissues, and the brain is identified as high-risk organs. For the analysis of other organs, cells from the hippocampus have low expression level of TMPRSS2 and ACE2, and the cerebellum, spinal cord, and neuronal epithelium data show zero expression of TMPRSS2 and ACE2 (Supplementary File: Figures S23-S26), which demonstrates a low infection risk of these organs.

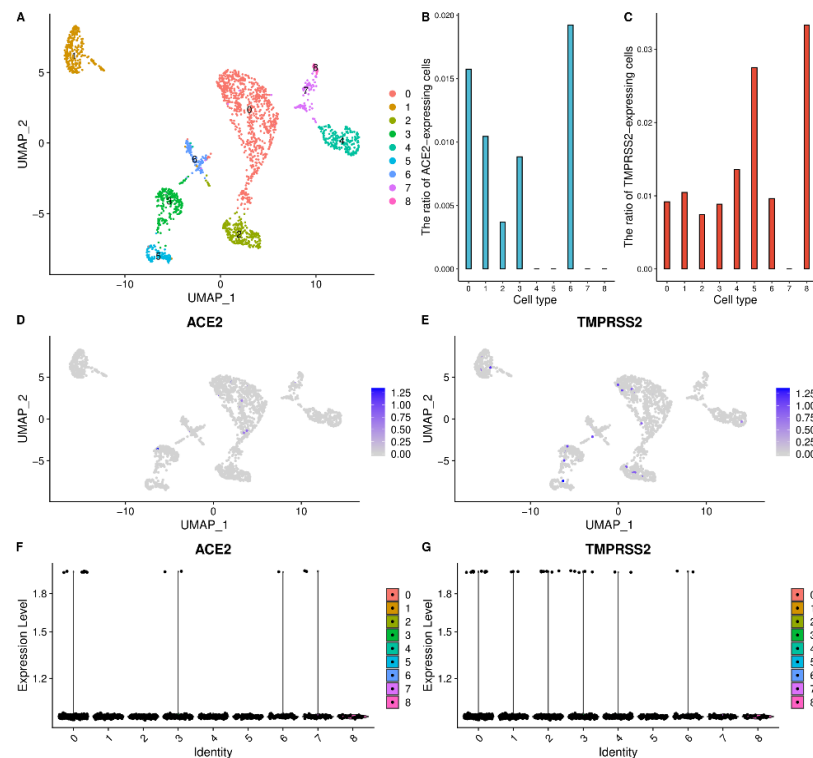


Fig. 3 High ACE2 and TMPRSS2 expression level of oligodendrocyte precursor cells and astrocytes in the substantia nigra and cortex. **a)** UMAP visualization of clustering results for the substantia nigra and cortex cells. **b)** The ratio of ACE2-expressed cells in each cell cluster. **c)** The ratio of TMPRSS2-expressed cells in each cell cluster. **d)** ACE2 expression level in each cell cluster on the UMAP plot. **e)** TMPRSS2 expression level in each cell cluster on the UMAP plot. **f)** The expression distribution of ACE2 across each cell cluster. **g)** The expression distribution of TMPRSS2 across each cell cluster.

Table 4: Cell types with high expression levels of ACE2 and TMPRSS2 in nervous system

Organs	Cell type	Ratio of ACE2-expressed cells	Ratio of TMPRSS2-expressed cells
Brain (Substantia nigra and cortex)	Oligodendrocyte precursor cell	1 6%	0 9%
	Astrocyte	1 9%	1 0%

Reproductive system

The scRNA-seq data of the testis, fallopian tube, ovary, and uterus from the reproductive system were collected for analysis. The ratios of the TMPRSS2-expressed cell and the ACE2-expressed cell in the epithelial cells of the fallopian tube are 26 5% and 1 4% respectively (Fig. 4, Table 5), and the ovarian somatic cells contain 1% TMPRSS2-expressed cells and 1% ACE2-expressed cells, so the fallopian tube is identified as a high-risk organ. The testis is also identified as a high-risk organ because of the high expression level of TMPRSS2 and ACE2 (Supplementary File: Figure S27, Table 5). Low ratios of ACE2-expressed cells in the ovary and uterus mean the ovary and uterus are low infection risk organs (Supplementary File: Figures S28-S29).

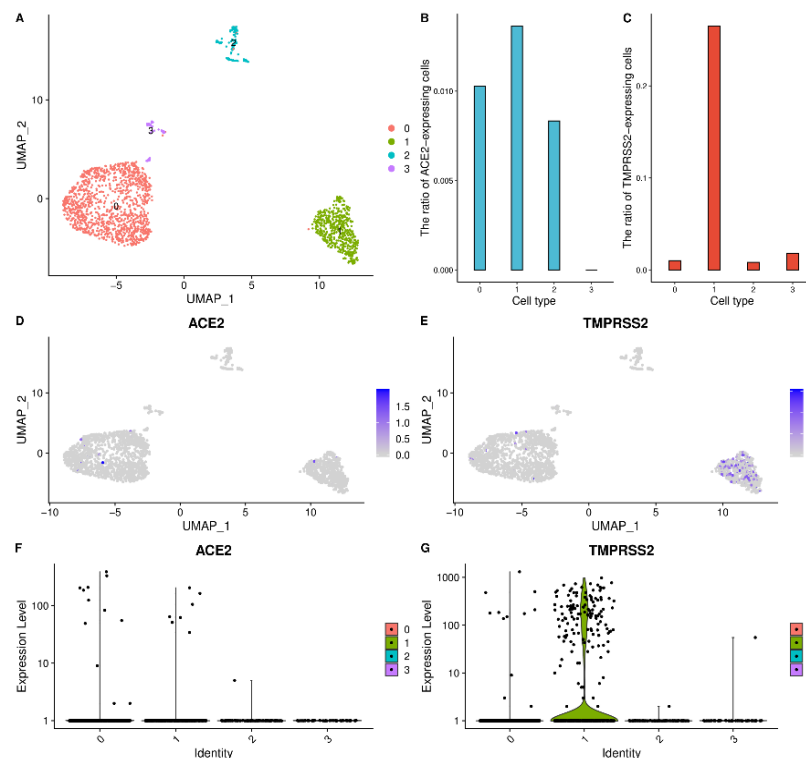


Fig. 4 High ACE2 and TMPRSS2 expression level of epithelial cells and ovarian somatic cells in the fallopian tube. a) UMAP visualization of clustering results for the fallopian tube cells. b) The ratio of ACE2-expressed cells in each cell cluster. c) The ratio of TMPRSS2-expressed cells in each cell cluster. d) ACE2 expression level in each cell cluster on the UMAP plot. e) TMPRSS2 expression level in each cell cluster on the UMAP plot. f) The expression distribution of ACE2 across each cell cluster. g) The expression distribution of TMPRSS2 across each cell cluster.

Table 5: Cell types with high expression levels of ACE2 and TMPRSS2 in reproductive system

Organs	Cell type	Ratio of ACE2-expressed cells	Ratio of TMPRSS2-expressed cells
Fallopian tube	Epithelial cells	1.4%	26.5%
	Ovarian somatic cell	1.0%	1.0%
Testis	Spermatogonium	1.7%	4.5%
	Peritubular myoid cell	1.7%	6.3%
	Testis somatic cell	2.1%	4.3%
	Spermatogonial stem cell	1.4%	2.1%

Circulatory system

The data of the heart, spleen, and artery from the circulatory system were collected to infer the susceptibility of these organs. The cardiomyocytes and cardiovascular progenitor cells from heart contain 6.6% and 12.5% ACE2-expressed cells respectively, and the TMPRSS2 is expressed in both cell clusters as well. Consequently, the heart is considered as a high-risk organ (Supplementary File: Figure S30, Table 6). Nevertheless, almost no cells of the spleen, artery, and peripheral blood data show high TMPRSS2 and ACE2 expression levels, which means they are low-risk organs (Supplementary File: Figures S31-S33).

Table 6: Cell types with high expression levels of ACE2 and TMPRSS2 in circulatory system

Organs	Cell type	Ratio of ACE2-expressed cells	Ratio of TMPRSS2-expressed cells
Heart	Cardiomyocyte	6.6%	0.8%
	Cardiovascular progenitor cell	12.5%	0.4%

Urinary System

The scRNA-seq data of the kidney, ureter, and prostate from the urinary system were utilized for the data analysis. The analysis results of the kidney scRNA-seq data show high ACE2 and TMPRSS2 expression levels in the nephron epithelial cells, epithelial cells, endothelial cells, and mesangial cells. Particularly, the ratios of TMPRSS2-expressed are 10.7%, 9.6%, 12.8%, and 14.5% respectively, and the ratios of ACE2-expressed are 2.7%, 2.7%, 2.7%, and 3.0% respectively (Supplementary File: Figure S34, Table 7). Therefore, the kidney is considered as high-risk organs. In addition, the ACE2 is not expressed in the ureter and prostate cells (Supplementary File: Figures S35-S36), and they are predicted as low-risk infection organs.

Table 7: Cell types with high expression levels of ACE2 and TMPRSS2 in urinary system

Organs	Cell type	Ratio of ACE2-expressed cells	Ratio of TMPRSS2-expressed cells
Kidney	Nephron epithelial cell	2.7%	10.7%
	Epithelial cell	2.7%	9.6%
	Endothelial cell	2.7%	12.8%
	Mesangial cell	3.0%	14.5%

Endocrine system, Motor system and Immune system

The susceptibility of organs from the endocrine system, immune system, and motor system was also concerned. In the endocrine system, almost no cells from the thyroid gland data show high ACE2 and TMPRSS2 expression levels, and the thymus gland data contains no ACE2-expressed cells (Supplementary File: Figures S37-S38). Hence, they are not high-risk organs. Likewise, few ACE2-expressed cells are found in the muscle from the motor system (Supplementary File: Figure S39). There is no ACE2 expression in the lymph nodes, tonsil (tonsil dendritic cells), and bone marrow data from the immune system (Supplementary File: Figures S40-S42), which mean they are low-risk organs.

The risk levels of susceptible organs

Based on TMPRSS2 expression level, we grouped the susceptible organs into three risk levels. In table 8, it is shown that lung, large intestine (colon and rectum), fallopian tube, and nose (nasal airway epithelium) are the most susceptible organs with TMPRSS2-expressed ratio over 20%, and the result indicates the SARS-CoV-2 mainly attacks the respiratory system, the digestive system, and the reproductive system. The kidney, Small intestine (duodenum and jejunum), and testis are susceptible organs with moderate risk. In addition, the esophagus, gall bladder, brain (substantia nigra and cortex), and heart are identified to be the potentially susceptible organs.

Table 8: Risk level of organs infection by SARS-CoV-2

Organs	Systems	Ratios of TMPRSS2-expressed cells	Cell types	Risk levels
Lung	Respiratory system	0.212	AT2	1
Large intestine (Rectum)	Digestive system	0.873	Goblet cell	1
Large intestine (Colon)	Digestive system	0.524	Goblet cell	1
Fallopian tube	Reproductive system	0.265	Epithelial cell	1
Nose (Nasal airway epithelium)	Respiratory system	0.234	All cells	1
Nose (Nasal brushing epithelial cells)	Respiratory system	0.188	Plasma cell	2
Kidney	Urinary system	0.145	Mesangial cell	2
Nose (Nasal turbinate epithelial cells)	Respiratory system	0.088	Plasma cell	2
Small intestine (Duodenum)	Digestive system	0.068	Enterocyte progenitor cell	2
Small intestine (Jejunum)	Digestive system	0.066	Enterocyte progenitor cell	2
Testis	Reproductive system	0.063	Peritubular myoid cell	2
Esophagus	Digestive system	0.049	Secretory progenitor cell	3
Small intestine (Ileum)	Digestive system	0.042	Intestinal epithelial stem cell	3
Gall bladder	Digestive system	0.026	Primordium cell	3
Brain (Substantia nigra and cortex)	Nervous system	0.010	Astrocyte	3
Heart	Circulatory system	0.008	Cardiomyocyte	3

Discussion

The clinical symptoms of patients infected by COVID are mainly manifested in the respiratory system and digestive system, including cough, shortness of breath, dyspnea, and diarrhea. However, some patients also developed symptoms such as heart damage and kidney failure, indicating that the virus affected the normal function of the circulatory and urinary systems. We investigated the susceptibility of the organs and tissues in various human systems based on the scRNA-seq data analysis. In details, 31 organs from nine major human systems were considered, in which 11 organs are identified to be susceptible to the virus. Moreover, we classified these susceptible organs into three levels in terms of their risk, which provide novel ideas for the follow-up detection of virus, treatment, and the monitoring of recrudescence.

However, due to the limitation of the data, only partial organs or tissues were involved, so the susceptible organs may be missed since the mechanism of cells infected by the COVID-19 is still not completely known. Furthermore, these organs reported to be susceptibility by the scRNA-seq data, should be further confirmed by the clinical observation and biological experiments.

Authors' contributions

Shuilin Jin and Jing Qi conceived and designed the study. Jiao Hua and Liying Zhang performed the data analysis. Yang Zhou, Jianlin Bian, and Beibei Liu collected for the available data. Jing Qi and Yang Zhou wrote the paper. Shuilin Jin, Jing Qi, Yang Zhou, Jiao Hua, Liying Zhang, Jialin Bian, Beibei Liu, and Zicen Zhao revised the manuscript. All authors read and approved the final manuscript.

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References

- 1 Gorbalenya AE, Baker SC, Baric RS, et al. Severe acute respiratory syndrome-related coronavirus: The species and its viruses – a statement of the Coronavirus Study Group. bioRxiv 2020; published online February 7. DOI:10.1101/2020.02.07.937862 (preprint).
- 2 Chen N, Zhou M, Dong X, et al. Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. Lancet 2020; **395**: 507–13.
- 3 Huang C, Wang Y, Li X, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. Lancet 2020; **395**: 497–506.
- 4 He L, Ding Y, Zhang Q, et al. Expression of elevated levels of pro-inflammatory cytokines in SARS-CoV-infected ACE2+ cells in SARS patients: relation to the acute lung injury and pathogenesis of SARS. The Journal of Pathology 2006; **210**: 288–97.
- 5 Li W, Sui J, Huang I, et al. The S proteins of human coronavirus NL63 and severe acute respiratory syndrome coronavirus bind overlapping regions of ACE2. Virology 2007; **367**: 367–374.
- 6 Wu K, Li W, Peng G, et al. Crystal structure of NL63 respiratory coronavirus receptor-binding domain complexed with its human receptor. Proceedings of the National Academy of Sciences of the United States of America 2009; **106**: 19970–74.
- 7 Ou X, Liu Y, Lei X, et al. Characterization of spike glycoprotein of SARS-CoV-2 on virus entry and its immune cross-reactivity with SARS-CoV. Nature Communications 2020; **11**: 1620.
- 8 Hoffmann M, Kleineweber H, Schroeder S, et al. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. Cell 2020; publish online March 5. DOI: 10.1016/j.cell.2020.02.052.
- 9 He Q, Mok T, Yun L, et al. Single cell RNA sequencing analysis of human kidney reveals the presence of ACE2 receptor: a potential pathway of COVID-19 infection. SSRN 2020; published online March 2. <http://dx.doi.org/10.2139/ssrn.3544810> (preprint).
- 10 Gallagher TM, Buchmeier MJ. Coronavirus spike proteins in viral entry and pathogenesis. Virology 2001; **279**: 371–74.
- 11 Meng T, Cao H, Zhang H, et al. The insert sequence in SARS-CoV-2 enhances spike protein cleavage by TMPRSS. bioRxiv 2020; published online February 8. DOI:10.1101/2020.02.08.926006 (preprint).
- 12 Wan Y, Shang J, Graham R, et al. Receptor recognition by the novel coronavirus from Wuhan: an analysis based on decade-long structural studies of SARS. Journal of Virology 2020; **94**.
- 13 Liu Q, Liu L, Ren L, et al. General observations of the cadaver system autopsy of the deceased

- COVID-19. *Journal of Forensic Medicine* 2020; **36**: 21–23.
- 14 Zhao Y, Zhao Z, Wang Y, et al. Single-cell RNA expression profiling of ACE2, the putative receptor of Wuhan COVID-19. *bioRxiv* 2020; published online January 26. DOI:10.1101/2020.01.26.919985 (preprint).
- 15 Wu C, Zheng S, Chen Y, et al. Single-cell RNA expression profiling of ACE2, the putative receptor of Wuhan 2019-nCoV, in the nasal tissue. *medRxiv* 2020; published online February 11. DOI:10.1101/2020.02.11.20022228 (preprint).
- 16 Wu C, Zheng M. Single-cell RNA expression profiling shows that ACE2, the putative receptor of Wuhan 2019-nCoV, has significant expression in the nasal, mouth, lung and colon tissues, and tends to be co-expressed with HLA-DRB1 in the four tissues. *Preprints* 2020; published online February 7. 2020020247 (preprint).
- 17 Zou X, Chen K, Zou J, et al. Single-cell RNA-seq data analysis on the receptor ACE2 expression reveals the potential risk of different human organs vulnerable to Wuhan 2019-nCoV infection. *Frontiers of Medicine* 2020; published online March 12. DOI:10.1007/s11684-020-0754-0.
- 18 Pan XW, Xu D, Zhang H, et al. Identification of a potential mechanism of acute kidney injury during the COVID-19 outbreak: a study based on single-cell transcriptome analysis. *Intensive Care Medicine* 2020; published online March 20. DOI:10.1007/s00134-020-06026-1.
- 19 Lin W, Hu L, Zhang Y, et al. Single-cell analysis of ACE2 expression in human kidneys and bladders reveals a potential route of 2019-nCoV infection. *bioRxiv* 2020; published online February 8. DOI:10.1101/2020.02.08.939892 (preprint).
- 20 Zhang H, Kang Z, Gong H, et al. The digestive system is a potential route of 2019-nCoV infection: a bioinformatics analysis based on single-cell transcriptomes. *bioRxiv* 2020; published online January 30. DOI:10.1101/2020.01.30.927806 (preprint).
- 21 Wang Z, Xu X. scRNA-seq profiling of human testes reveals the presence of ACE2 receptor, a target for SARS-CoV-2 infection, in spermatogonia, leydig and sertoli cells. *Preprints* 2020; published online February 21. DOI:10.20944/preprints202002.0299.v1 (preprint).
- 22 Zhang J, Wu Y, Wang R, et al. Bioinformatic analysis reveals that the reproductive system is potentially at risk from SARS-CoV-2. *Preprints* 2020; published online February 21. DOI:10.20944/preprints202002.0307.v1 (preprint).
- 23 Liu F, Long X, Zou W, et al. Highly ACE2 expression in pancreas may cause pancreas damage after SARS-CoV-2 infection. *medRxiv* 2020; published online February 28. DOI:10.1101/2020.02.28.20029181 (preprint).
- 24 Madissoon E, Wilbreyclark AL, Miragaia RJ, et al. Lung, spleen and oesophagus tissue remains stable for scRNAseq in cold preservation. *bioRxiv* 2019; published online August 31. DOI:10.1101/741405.
- 25 Han XP, Zhou ZM, Fei LJ, et al. Construction of a human cell landscape at single-cell level. *Nature* 2020; published online March 25. DOI: 10.1038/s41586-020-2157-4.
- 26 Gutierrez GD, Kim J, Lee A, et al. Gene signature of the human pancreatic ϵ -cell. *Endocrinology* 2018; **159**: 4023–32.
- 27 Onorati M, Li Z, Liu F, et al. Zika virus disrupts phospho-TBK1 localization and mitosis in human neuroepithelial stem cells and radial glia. *Cell Reports* 2016; **16**: 2576–92.
- 28 Zhong S, Ding W, Sun L, et al. Decoding the development of the human hippocampus. *Nature* 2020; **577**: 531–36.
- 29 Fan X, Bialecka M, Moustakas I, et al. Single-cell reconstruction of follicular remodeling in the

- human adult ovary. *Nature Communications* 2019; **10**: 3164.
- 30 Hu Z, Artibani M, Alsaadi A, et al. The repertoire of serous ovarian cancer non-genetic heterogeneity revealed by single-cell sequencing of normal fallopian tube epithelial cells. *Cancer Cell* 2020; **37**: 226–242.e7.
- 31 Guo J, Grow EJ, Mlcochova H, et al. The adult human testis transcriptional cell atlas. *Cell Research* 2018; **28**: 1141–57.
- 32 Garcia SR, Deprez M, Lebrigand K, et al. Novel dynamics of human mucociliary differentiation revealed by single-cell RNA sequencing of nasal epithelial cultures. *Development* 2019; **146**: dev177428.
- 33 Reyfman PA, Walter JM, Joshi N, et al. Single-cell transcriptomic analysis of human lung provides insights into the pathobiology of pulmonary fibrosis. *American Journal of Respiratory and Critical Care Medicine* 2019; **199**: 1517–36.
- 34 Cui Y, Zheng Y, Liu X, et al. Single-cell transcriptome analysis maps the developmental track of the human heart. *Cell Reports* 2019; **26**: 1934–50.e5.
- 35 Tanghuau T, Gueguen P, Goudot C, et al. Human in vivo-generated monocyte-derived dendritic cells and macrophages cross-present antigens through a vacuolar pathway. *Nature Communications* 2018; **9**: 2570.
- 36 Takeda A, Hollmen M, Dermadi D, et al. Single-cell survey of human lymphatics unveils marked endothelial cell heterogeneity and mechanisms of homing for neutrophils. *Immunity* 2019; **51**: 561–72.e5.